# Student Computing Club: <br> Dimension reduction algorithms for visualizing single-cell genomic data using R 

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Motivating example

## Dimension reduction



## population

- Basophils
- CD16-_NK_cells

CD16+_NK_cells CD34+CD38+CD123-_HSPCs

- CD34+CD38+CD123+_HSPCs
- CD34+CD381o_HSCs
- CD4_T_cells

CD8_T_cells

- Mature_B_cells
- Monocytes
- pDCs
- Plasma_B_cells
- Pre_B_cells
- Pro B cells unassigned


## Single-cell data

## Single-cell data

Example: Mass cytometry (CyTOF)

targeted set of proteins of interest; bind to known antibodies

## Single-cell data

Example: Single-cell RNA sequencing (scRNA-seq)


## Dimension reduction

## Dimension reduction

Issue: too many dimensions!

How to represent visually?
$\rightarrow$ exploratory data analysis; presentation of results (reveal or display patterns of interest, e.g. clusters, trajectories, differential sample features)

How to analyze computationally?
$\rightarrow$ curse of dimensionality; computational scalability

## Dimension reduction

Summarize data using a lower number of dimensions

Single-cell data: two main applications

- visualization (i.e. plot in 2 or 3 dimensions)
- data preprocessing (curse of dimensionality, remove noise, correlated features, computational scalability)

Dimension reduction algorithms

- select or calculate smaller number of dimensions (features) that capture the underlying patterns of interest in the dataset
- many approaches
- relevant patterns depend on scientific question


## Examples

## Dataset

Levine_32dim: mass cytometry (CyTOF) dataset from Levine et al. (2015)

- healthy human bone marrow mononuclear cells (BMMCs)
- 32 surface protein markers
- reference cell population (cluster) labels for 14 immune cell populations
- 265,627 cells (104,184 or 39\% assigned)
- previously used to benchmark clustering algorithms in our publication (Weber and Robinson, 2016); available as formatted R/Bioconductor objects via HDCytoData package (Weber and Soneson, 2019)


## Example: principal component analysis (PCA)

Intuitively: sequentially project data onto rotated orthogonal axes, where each axis captures maximal amount of remaining variance in data

Linear algorithm

- reduced dimensions (principal components) can be interpreted as combinations of original dimensions

Single-cell data

- PCA commonly used for preprocessing, i.e. reduce dimensionality prior to downstream analysis (e.g. keep top 50 or 100 PCs in scRNA-seq data)
- Often does not work well for visualization, due to nonlinear data structure


## Example: principal component analysis (PCA)

Levine_32dim dataset

population

- Basophils
- CD16-_NK_cells CD16+_NK_cells CD34+CD38+CD123-_HSPCs
- CD34+CD38+CD123+_HSPCs
- CD34+CD381o_HSCs
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CD8_T_cells

- Mature_B_cells
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- pDCs
- Plasma_B_cells
- Pre_B_cells
- Pro_B_cells unassigned


## Example: t-SNE

t-Distributed Stochastic Neighbor Embedding (t-SNE) (van der Maaten and Hinton, 2008; van der Maaten 2014)

Developed for visualizing datasets in machine learning; quickly adopted by single-cell biology community

Nonlinear algorithm

Single-cell data

- Advantages: tends to clearly separate clusters (cell populations)
- Disadvantages: reduced dimensions difficult to interpret (especially global distances); can "force" cluster structure; computational scalability


## Example: t-SNE

Levine_32dim dataset


## population

- Basophils
- CD16-_NK_cells

CD16+_NK_cells
CD34+CD38+CD123-_HSPCs

- CD34+CD38+CD123+_HSPCs
- CD34+CD38lo_HSCs
- CD4_T_cells
- CD8 T cells
- Mature B cells
- Monocytes
- pDCs
- Plasma_B_cells
- Pre B cells
- Pro_B_cells
unassigned


## Example: UMAP

Uniform Manifold Approximation and Projection (UMAP) (McInnes et al. 2018)

Widely adopted for single-cell data within the last year

Nonlinear algorithm

Single-cell data

- Advantages: tends to separate clusters as well as t-SNE but preserves global distances more accurately; computationally efficient


## Example: UMAP

Levine_32dim dataset

population

- Basophils
- CD16-_NK_cells

CD16+_NK_cells
CD34+CD38+CD123-_HSPCs
CD34+CD38+CD123+_HSPCs

- CD34+CD38lo_HSCs
- CD4_T_cells

CD8_T_cells

- Mature_B_cells
- Monocytes
- pDCs
- Plasma_B_cells
- Pre_B_cells
- Pro_B_cells
unassigned

More examples

## Results

Reduced dimension plots for each method/dataset: Samusik_01 dataset (CyTOF)



labels
B-cell Frac A-C (pro-B cells)
Basophils
CD4 T cells
CD8 T cells
Classical Monocytes
CLP
CMP
Eosinophils
gd T cells
GMP
HSC
IgD- IgMpos B cells
IgDpos IgMpos B cells
$\operatorname{lgM}$ - $\operatorname{lgD}$ - B-cells Intermediate Monocytes Macrophages mDCs
MEP
MPP

- NK cells

NKT
Non-Classical Monocytes pDCs Plasma Cells unassigned

## Results

Reduced dimension plots for each method/dataset: Koh dataset (scRNA-seq)




## Results

Reduced dimension plots for each method/dataset: Trapnell dataset (scRNA-seq)




## Interactive demo

# Interactive demo 

See RStudio

Thank you!

Additional slides

## Curse of dimensionality

Standard (e.g. Euclidean) distances become largely meaningless in very high-dimensional spaces
$\rightarrow$ all points reside in thin "shell" of high-dimensional sphere or cube, with ~zero interior volume; all points are approximately the same distance apart

|  | 2-dimensional | $\varepsilon=0.01$ |  |  |
| :---: | :---: | :---: | :---: | :---: |
| 1-dimensional |  | $p$ | $(1-\varepsilon)^{p}$ | $1-(1-\varepsilon)^{p}$ |
|  | $1-(1-\varepsilon)^{2}$ | 1 | 0.99 | 0.01 |
| H   <br> $\mathrm{H} / 2$ $1-\varepsilon$ $\mathrm{e} / 2$ | \$ $\geqslant \leqslant \geqslant \leqslant \geqslant$ | 2 | 0.9801 | 0.0199 |
|  |  | 3 | 0.9703 | 0.0297 |
|  | * $(1-\varepsilon)^{2}$ | $\ldots$ |  |  |
|  | * | 10 | 0.9044 | 0.0956 |
|  | - | 100 | 0.3660 | 0.6340 |
|  |  | 1000 | 4.32e-05 | ~1.0 |
|  |  | 10000 | $2.25 \mathrm{e}-44$ | ~1.0 |

